

Association between two *TRPM2* mutations and Painful Small Fiber Neuropathy: from NGS analysis to functional characterization

Kaalindi Misra¹, Silvia Santoro¹, Andrea Barbieri^{2,3}, Margherita Marchi⁴, Núria Comes^{2,3}, Gerard Callejo^{2,3}, Erika Salvi⁴, Massimo Filippi^{5,6,7,8}, Xavier Gasull^{2,3}, Giuseppe Lauria^{4,9}, Federica Esposito^{1,5}

¹ Laboratory of Human Genetics of Neurological Disorders, IRCCS San Raffaele Scientific Institute, INSPE, Milan, (Italy); ² Neurophysiology Laboratory, Department of Biomedicine, Medical School, Institute of Neurosciences, Universitat de Barcelona, Barcelona, Spain; ³ Institut d'Investigacions Biomèdiques August Pi i Sunyer (IDIBAPS), Barcelona, Spain; ⁴ Neurology Unit, Foundation IRCCS Carlo Besta Neurological Institute, Milan (Italy); ⁵ Neurology and Neurorehabilitation Unit, IRCCS San Raffaele Scientific Institute, Milan (Italy); ⁶ Vita-Salute San Raffaele University, Milan (Italy); ⁷ Neurophysiology Unit, IRCCS San Raffaele Scientific Institute, Milan (Italy); ⁸ Neuroimaging Research Unit, Institute of Experimental Neurology, Division of Neuroscience, IRCCS San Raffaele Scientific Institute, Milan (Italy); ⁹ Department of Biomedical and Clinical Sciences "Luigi Sacco", University of Milan, Milan (Italy)

Introduction

- Small fiber neuropathy (SFN) is a heterogeneous group of disorders affecting thin myelinated A δ and unmyelinated C fibers
- Clinical symptoms include neuropathic pain, possible loss of thermal and nociceptive sensation, and autonomic disturbances [3]
- The aetiology of SFN is complex, involving both inherited and acquired factors
- Genetic studies in painful SFN revealed that Voltage Gated Sodium Channels (VGSC) genes, particularly *SCN9A*, are involved in pain amplification [1,2]
- Besides VGSC genes, Transient Receptor Potential (TRP) channel genes have also been associated with neuropathic pain because of their pivotal role in nociception [6]

Aim

To broaden genetic knowledge of painful SFN via whole-exome sequencing (WES) on Italian families with SFN and to investigate mutation impacts on channel function

Methods

- Twelve families with painful SFN were selected having at least one affected member, positive neurological examination and pain questionnaire result with numerical rating score ≥ 4 .
- After performing WES, Variants were filtered, keeping only the ones mapping to a manually curated panel of pain related genes (n=592), with minor allele frequency $\leq 5\%$ in population databases [4]; conservation and computational predictors were also considered in the filtering. Segregation analysis was performed in each pedigree.
- The impact of selected causative variants was assessed through electrophysiological patch-clamp recordings on HEK cell line cultures transfected with wildtype and mutant expression vectors [5].

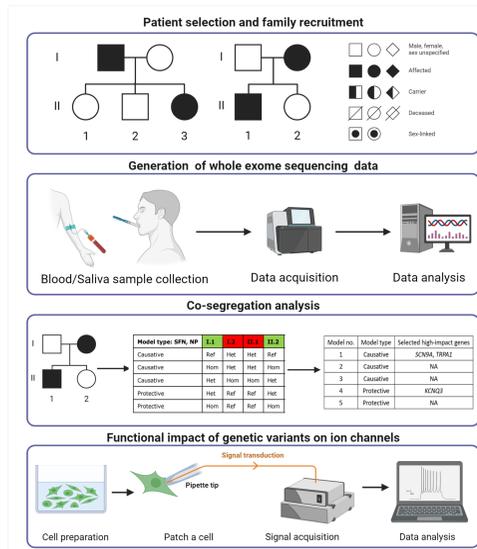


Figure 1: Graphical representation of methodology of the study

Results

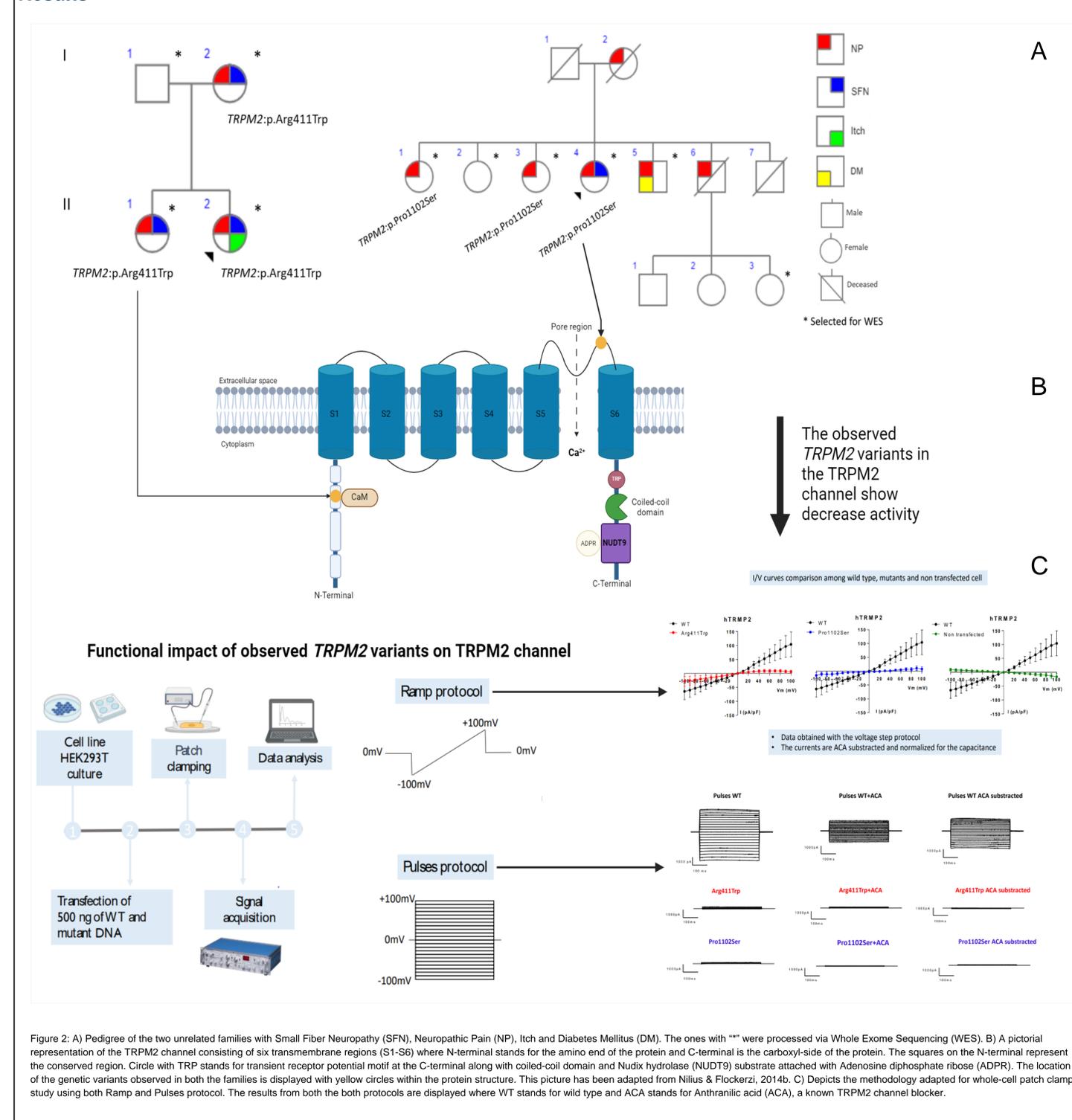


Figure 2: A) Pedigree of the two unrelated families with Small Fiber Neuropathy (SFN), Neuropathic Pain (NP), Itch and Diabetes Mellitus (DM). The ones with *** were processed via Whole Exome Sequencing (WES). B) A pictorial representation of the *TRPM2* channel consisting of six transmembrane regions (S1-S6) where N-terminal stands for the amino end of the protein and C-terminal is the carboxyl-side of the protein. The squares on the N-terminal represent the conserved region. Circle with TRP stands for transient receptor potential motif at the C-terminal along with coiled-coil domain and Nudix hydrolase (NUDT9) substrate attached with Adenosine diphosphate ribose (ADPR). The location of the genetic variants observed in both the families is displayed with yellow circles within the protein structure. This picture has been adapted from Nilius & Flockerzi, 2014b. C) Depicts the methodology adapted for whole-cell patch clamp study using both Ramp and Pulses protocol. The results from both the both protocols are displayed where WT stands for wild type and ACA stands for Anthranilic acid (ACA), a known *TRPM2* channel blocker.

Conclusion

- This study reports **two novel missense mutations in *TRPM2*** segregating with painful SFN.
- Functional analysis revealed that both mutations affect the normal function of the *TRPM2* channel, leading to a significant reduction in their currents.
- TRPM2* channel was previously linked to neuropathic pain in animal models but not in human genetic studies on neuropathic pain or SFN.
- These findings suggest that these mutations are likely causative and contribute to painful SFN development.
- Further investigations are warranted to elucidate the precise mechanisms by which these mutations lead to painful SFN and to speculate on *TRPM2* as a potential pharmacological druggable target in future studies.

Future Studies

- Calcium Imaging:** Measure intracellular calcium levels to assess *TRPM2* channel activity and calcium influx in mutant vs. wild-type channels.
- Immunocytochemistry:** Visualize the localization of *TRPM2* in cells to see if mutations alter its distribution.
- Co-immunoprecipitation:** Study interactions between *TRPM2* and other proteins, like calmodulin, to understand if mutations disrupt regulatory mechanisms.

References

- Brouwer BA, Merkies ISJ, Gerrits MM, Waxman SG, Hoelmakers JGJ, Faber CG. Painful neuropathies: the emerging role of sodium channelopathies. *J Peripher Nerv Syst* 2014;19:53-65. doi:10.1111/jn.512071.
- Han C, Hoelmakers JGJ, Liu S, Gerrits MM, Te Morsche RHM, Lauria G, De-Haj SD, Dierich JPH, Faber CG, Merkies ISJ, Waxman SG. Functional profiles of *SCN9A* variants in dorsal root ganglion neurons and superior cervical ganglion neurons correlate with autonomic symptoms in small fibre neuropathy. *Brain* 2012;135:2613-2628. doi:10.1093/brain/aww187.
- Hovagimian A. Gibbins CH. Diagnosis and treatment of pain in small-fiber neuropathy. *Curr Pain Headache Rep* 2011;15:193-200. doi:10.1007/s11916-011-0181-7.
- Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Akioti J, Wang Q, Collins RL, Laricchia KM, Ganna A, Bierbaum DP, Gauthier JD, Brandt H, Solominski M, Walters NA, Rohrer D, Singarayer M, England EM, Soyak EG, Kosmicki JA, Walters RK, Tashman K, Farjuna Y, Banks E, Poterba T, Wang A, Sedo C, Whiffin N, Chong JX, Samocha KE, Pierce-Hoffman E, Zappala Z, O'Donnell-Luria AH, Minkkel EV, Weisburd B, Lek M, Ware JS, Vitell C, Armean IM, Bergelson L, Cibulskis K, Connolly KM, Covarrubias M, Donnelly S, Farnham S, Gauthier J, Gupta N, Jangdevi T, Kaplan D, Liawarska C, Manrai R, Neveval S, Petrillo N, Roazen D, Ruano-Rubio V, Saltzman A, Schlicher M, Soto J, Tibbetts K, Tolonen C, Wade G, Talkowski ME, Agular Salinas CA, Ahmad T, Albert CM, Ardissino D, Atzmon G, Barnard J, Beaugerie L, Benjamin EJ, Boehnke M, Bonnycastle LL, Botttinger EP, Bowden DWJ, Brown MJ, Chambers JC, Chan JC, Chasman D, Cho J, Chung MK, Cohen B, Conroy A, Dabelea D, Daly MJ, Darbar D, Duggirala R, Dupuis J, Ellorin PT, Eskova R, Erdmann J, Esko T, Färkkilä M, Florez J, Franke A, Freitag G, Glaser B, Glan S, Gokhale S, Gonzalez C, Groop L, Haanen C, Harris C, Harms M, Hekunen M, Heli MM, Hultman CM, Kallala M, Kaprio J, Kathiresan S, Kim BJ, Kim YJ, Kiryl G, Kooper J, Koskinen S, Krumholz HM, Kugathasan S, Kwak SH, Laakso M, Lehtimäki T, Loos RFJ, Lubitz SA, Ma RCW, MacArthur DG, Marrugat J, Mattila KM, McCarron SE, McCarthy MI, McGovern D, McPherson R, Meigs JB, Melander O, Metspalu A, Neale BM, Nilsson PM, O'Donovan MC, Ongur D, Orozco L, Owen MJ, Palmer CNA, Palanisami A, Park KS, Patil C, Palver AE, Rahaman N, Remes AM, Rioux JD, Ripatti S, Roden DM, Saleheen D, Salonen V, Samani NJ, Scharf J, Schunkert H, Shomekhar MB, Sklar P, Sorainen H, Sorokina H, Spector T, Sullivan PF, Suvisaari J, Tai ES, Teo YY, Tiihonen M, Tuomari M, Turner D, Tusie-Luna T, Vartiainen E, Watkins H, Weersma RK, Weisman M, Wilson JG, Xavier RJ, Yeele BM, Daly MJ, MacArthur DG. The mutational constraint spectrum quantified from variation in 141,456 humans. *Nat* 2020;581:809-2020:581:434-443. doi:10.1038/s41586-020-2308-7.
- Lev S, Mink B. Constitutive Activity of TRP Channels: Methods for Measuring the Activity and Its Outcome. *Methods Enzymol* 2010;484:591-612.
- Ślęczkowska M, Misra K, Santoro S, Gerrits MM, Hoelmakers JGJ. Ion Channel Genes in Painful Neuropathies. *Biomedicines* 2023;11. doi:10.3390/biomedicines11102680.